# Study of the Impact of Heavy Metal (Cd, Pb and Hg) Contamination in Wedung Waters, Demak on Anadara granosa (Linnaeus, 1758)

#### Wahyu Andre Wijaya, Bambang Yulianto, and Wilis Ari Setyati

Department of Marine Sciences, Faculty of Fisheries and Marine Sciences, Diponegoro University, Jl. Prof. H. Soedarto, S.H, Tembalang, Semarang, Indonesia 50275.

Corresponding Author

#### **Abstract**

Wedung Waters, Demak in Central Java Indonesia is a potential location for harvesting Blood Cockles, Anadara granosa. Anthropogenic activities such as agriculture and fishing have potential to cause the entry of heavy metals like Cd, Pb, and Hg into marine environment and affect the biota A. granosa. The test of Cd, Pb, and Hg contaminations in waters, sediments and A. granosa soft tissues have been performed in this study. Histopathology analysis in gill and hepatopancreas are done to see the impact of the contaminations in that biota. The result revealed that Cd, Pb and Hg are not detected in waters, but two of the pollutions, Cd and Pb, are found in sediment and A. granosa soft tissues. Cd and Pb present about 0.08 - 0,2 mg.kg<sup>-1</sup> and 3,35-6,46 mg.kg<sup>-1</sup> respectively in sediment Heavy metal test revealed that the Cd and Pb content in soft tissue of A. granosa present about 0,56-0,70 mg.kg<sup>-1</sup> and 0,05 -0,1 mg.kg<sup>-1</sup> respectively. We found inflammation in gills of A. granosa that can be seen from the infiltration of hemocyte, hyperplasia of goblet cell, gill swelling, hemocyte granulocyte, pigmentation, and cilia degeneration. In digestive gland we found the Lifting of epithelial cell, infiltration of hemocyte, pigmentation, necrosis of epithelial cell.

**Keyword:** Anadara granosa, Heavy metal Cd,Pb,Hg, Histopathology

#### **INTRODUCTIONS**

Pollution contamination such as heavy metals from anthropogenic activities into the marine environment has been reported to increase in recent decades [2]. Heavy metals are natural elements of waters and sea waters. Anthropogenic activities like Industrial, agricultural runoff (pesticides, fertilizer and herbicides), acid rain etc. contributed to increase metal content in sediment or marine environment [2;9;20].

Heavy metal pollutant is one of the biggest problems that pose a threat to the ecosystem. Heavy metals have the potential for bioaccumulation and biomagnification that cause heavier exposure to some organisms [11]. Heavy metals toxic is presisten because it

has a tendency to accumulate in organisms and undergo food chain amplification, and the fact that they are not degraded make it even worse [38]. Accumulation of heavy metal in sediment can cause ecotoxicological effect in aquatic organism [14]. Heavy metal such Cd, Pb, and Hg have a high toxicity even in low doses and have the potential for biomagnification, bioaccumulation, and incorporation [45;40].

Bivalve is one of biota which is potential to be bio indicator of heavy metal contamination, especially for Cd and Pb [7]. *A. granosa* are recommended to be biomonitoring of heavy metal because of their capability to accumulate it. In Indonesia, *A. granosa* is a fairly commercial shellfish commodity. Production of shellfish in Indonesia from 2011 to 2015 increased by an average of 6.81% per year. In 2015, Indonesian shellfish production reached 59,613 tons where 85.26% of the number was from *A. granosa* [18]. Wedung is a sub-district in Demak Regency that contributes to the production of *A. granosa* species in Central Java Province of Indonesia [4].

Histopathology of gills and hepatopancreas is one of the tools used to view evidence of biomonitoring [17]. Some researchers see histopathological results on gills and hepatopancreas to be associated with the impact of heavy metal contamination on bivalves [32; 22; 37].

The aim of this study is to analyze Cd, Pb, and Hg in Wedung Waters, Demak in Central Java Indonesia and their effects on the bivalve of *A. granosa* by using histopathology analysis to the gills and digestive glands.

#### **MATERIALS AND METHODS**

The research was held in February and March 2019 at Wedung Waters, Demak Central Java Indonesia. The purposive sampling was done at four stations: Station A at 6°44'39.20"S - 110°33'4.37"E, Station B at 6°44'57.29"S - 110°33'17.02"E, Station C at 6°46'1.84"S - 110°32'21.60"E and Station D at 6°47'14.36"S - 110°33'25.79"E. The sampling locations are showed at Figure 1 below.

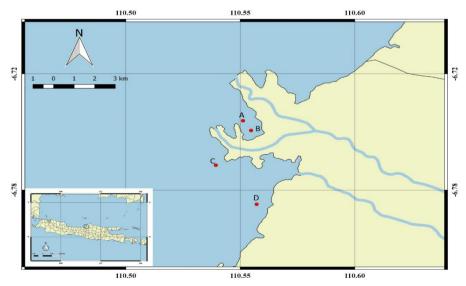


Figure 1. Sampling Location

Samples of 100 Blood Cockles catch using nets from each stations in February and March. The samples were stored in polyethylene plastic bags and put into a cool box. The temperature of the cool box were controlled to be below 4°C during transportation. Finally, the samples were stored in freezer with temperature of -40°C before heavy metal analysis [20].

500 ml of sea water sampling was taken using a *Nansen* bottle on a surface layer of water and stored in polyethylene bottle [34]. HNO<sub>3</sub> was added into the sample to get sample with pH <2 [25]. Sediment samples were taken by following the procedure in [16] at a depth of 0 to 5 cm using a portable Ekman grab sampler.

# **Water Parameter Quality Measurement**

Water parameter data such temperature, salinity, pH, and Dissolved Oxygen (DO) was taken directly /in situ during the study using Water Quality Checker [10;47]

## **Heavy Metal Analysis on Water and Sediment**

Heavy metals Cd, Pb, and Hg in water are analyzed by following the procedure in Indonesian National Standard [25] for (Cd) test, [27] for (Pb) test [26], and for (Hg) test. The test was carried out by homogenizing 50 ml of sample water for the Cd test and 100 ml of sample water for the Pb test, with 5 ml of concentrated nitric acid. The water sample then heated up until its decreased to 15 ml in volume. Next step was adding *aquadest* into the samples until it reached 50 ml in volume for the Cd test and 100 ml in volume for Pb test. The solution then analyzed by Atomic Absorption Spectroscopy (AAS) graphite type with a wavelength 228,8 nm for Cd test and 283,3 nm for Pb test.

The Hg analysis procedure for water was carried out by homogenizing 100 ml of water with H<sub>2</sub>SO<sub>4</sub> and 2.5 ml of HNO<sub>3</sub>. 15 ml of KMnO<sub>4</sub> solution was then added to the sample and waited for 15 minutes before adding 8 ml of K<sub>2</sub>S<sub>2</sub>O<sub>8</sub>. Samples were heated in a hot plate for 2 hours at 95°C before adding hydroxylamine-NaCl to reduce KMnO<sub>4</sub>. The final step was adding 5 ml of SnCl<sub>2</sub> before analyzing it using AAS hydric with a wavelength of 253.7 nm.

Grinded sediment sample using 100 meshes digested by weighed 0.1 g of a dry sample weighed beforehand carried out at around 170°C for 24 hours using a mixture of 2 ml of HNO<sub>3</sub>, 2 ml H<sub>2</sub>O<sub>2</sub> and 0.5 HF. After digestion, the solution filtrated and diluted to final volume of 25 ml to measured using ICP OES with wavelengths of 214,439 nm for Cd, 220,353nm for Pb and 184,887nm for Hg [33]

# **Heavy Metal Analysis on Blood Cockle**

Analysis of Cd and Pb on Blood Cockle refers to the Indonesian National Standard [23]. Amount of 5 grams of soft tissue of Blood Cockle being destructed at 450°C with a furnace for 18 hours. after that add 1 ml of 65% HNO<sub>3</sub> then evaporated on a hot plate until dry, sample destroyed again at 450°C for 3 hours, then added 5 ml of HCl 6 M and heated with a hot plate. After heating 10 ml of 0.1 M HNO<sub>3</sub> adding until the volume reaches 50 ml. Samples were analyzed by AAS with a wavelength, for Cd 228.8 nm, while Pb at 283.3 nm.

Hg test was carried out referring to the Indonesian National Standard SNI [24], a number of 5 grams of sample was added 3 pieces of boiling stone, then added 10 mg -  $20 \text{ mg V}_2\text{O}_5$  and given  $10 \text{ mL HNO}_3$  65% and  $10 \text{ mL H}_2\text{SO}_4$  95% - respectively 97%. The sample is then heated, then rinsed with 15 mL deionized water, 2 drops of 30%  $\text{H}_2\text{O}_2$  and finally added with distilled water to a volume of 100 ml. The heavy metal Hg in the sample was read using AAS with a wavelength of 253.7 nm.

## **Histopathology Analysis**

A. granosa was fixation using Davidson's solution then dehydrated with 70-95% and 100% ethanol, then planted using paraffin. The sample is then cut using a microtome. Furthermore, it was stained using Hematoxylin and Eosin. Histopathological results were read using a microscope with a magnification of 10x-1000x. [6].

#### RESULT AND DISCUSSION

#### **Water Parameter**

The temperature at the four station ranges from 28.8°C - 31.6°C which is common temperature in tropical waters. DO generally increases with temperature, DO ranges between 5.05- 5.4 mg l<sup>-1</sup>. pH ranges between 8 - 8.55 that make it possible for *A. granosa* to live. Salinity ranges between 16.70 - 29.11 ppt that is within the *A.granosa* habitat range [3]. Result of water parameter shown at Table 1.

Table 1. Water Parameter

Parameter		Month			
	A	В	C	D	
Temperature (°C)	$29,2 \pm 0,14$	29,35 ± 0,64	$30,4 \pm 0,00$	$31,55 \pm 0,49$	February
pН	$8 \pm 0,00$	$8,\!30\pm0,\!14$	$8,00 \pm 0$	$8,2\pm0,14$	
Salinity (ppt)	$18,9 \pm 0,14$	$18,15 \pm 0,02$	29,11 ± 1,12	$16,70 \pm 0,79$	
DO (mg.l <sup>-1</sup> )	$5,\!19\pm0,\!01$	$5,40 \pm 0,30$	$5,\!10\pm0,\!13$	$5,\!06 \pm 0,\!27$	
Temperature ( <sup>0</sup> C)	$28,8 \pm 0,00$	28,95 ± 0,07	29,85 ± 0,07	$31,6 \pm 0,14$	March
pН	$8,15 \pm 0.07$	$8,2 \pm 0,00$	$8,\!55 \pm 0,\!07$	$8,3 \pm 0,00$	
Salinity (ppt)	$20 \pm 0.16$	$18.56 \pm 0,002$	$26,445 \pm 0,49$	$17,265 \pm 1,75$	
DO (mg.l <sup>-1</sup> )	5,3 ± 0,21	5,135 ± 0,02	$5,055 \pm 0,09$	$5,24 \pm 0,37$	

## **Heavy Metal Test Results for Water and Sediments**

Heavy metal analysis of Cd, Pb, and Hg were not detected in seawater from four stations in Wedung Waters, Demak. The results showed that the lowest content of Cd in February was found at station C with a value of 0.08 mg.kg<sup>-1</sup> and the highest value at station D with a value of 0.11 mg.kg<sup>-1</sup>. While for sample that were taken in March 2019, the lowest value of the Cd found at station D with a value of 0.11 mg.kg<sup>-1</sup> and the highest at Station B and C with a value of 0.20 mg.kg<sup>-1</sup>. The lowest Pb content in February was found at Station A with a value of 3.35 mg.kg<sup>-1</sup> and the highest value at station D with a value of 5.38 mg.kg<sup>-1</sup>. The Pb content value in March was higher as compared to the previous month, with the lowest grade (Pb) value at station A of 4.77 mg.kg<sup>-1</sup> and the highest value at Station C with a value of 6.46 mg.kg<sup>-1</sup>. Hg content were not found at all stations both in February and March 2019. The complete test results are provided in Table 2.

**Table 2.** Content of Heavy Metals in Sediments

Parameter -	Heavy Metals content in Sedimen at Station (mg.kg <sup>-1</sup> )				Target Level	Month
	A	В	C	D	(mg.kg <sup>-1</sup> )*	WIOHUI
Cd	0,10 ±0,01	0,10 ±0	0,08 ±0	0,11 ±0	0.8	
Pb	$3,35 \pm 0,05$	$3,90\pm0,04$	$3,36 \pm 0,04$	$5,38 \pm 0,02$	85	February
Hg	Nd	Nd	Nd	Nd	0.03	
Cd	0.17 ±0.01	$0.20 \pm 0.02$	0.20 ±0.01	0.11 ±0.01	0.8	
Pb	$4.77 \pm 0.09$	$6.34 \pm 0.21$	$6.46 \pm 0.09$	$5.74 \pm 0.24$	85	March
Hg	Nd	Nd	Nd	Nd	0.03	

Source: Primary Data

\*IADC (International Association of Drilling Contractors) / CEDA Central Dredging Association) (1997)

Nd = Not detected

The Kruskal Wallis test of Cd content in sediments from four stations A, B, C, and D showed a *p-value* of 0.072 in February and 0.106 in March (> 0.05). Therefore, there was no significant difference of Cd content in sediment taken from the four stations. The Kruskal Wallis test for Pb levels in sediments resulted with *p-value* of 0.112 in February and 0.104 in March (> 0.05). That means there was no significant difference of Pb content in sediment taken from the four stations.

Hg content was not detected in the water and sediments. This indicates that the Wedung Waters are not polluted by Hg. Cd and Pb content were also not detected in the water but were detected in sediments. Cd and Pb content of sediments are below the threshold set by the CEDA [15] which is 0.8 mg.kg<sup>-1</sup> for Cd and 85 mg.kg<sup>-1</sup> for Pb. This means that Pb and Cd content in sediments in the Wedung Waters are not too dangerous for the environment.

By looking at the activities in Wedung, it is suspected that the contamination of Cd and Pb to marine environment comes from anthropogenic activities in the form of agriculture and vessel activities. Referring to the Statistics Data of the Wedung District (2015), the main livelihoods of the residents dominated by farmers with a total land area of 7.000,40 ha and fishermen with a total of 1,228 units of vessels. Agricultural activities in the form of the use of fertilizers, pesticides, and herbicides cause the contamination of Cd and Pb to the waters. [12] stated the presence of Cd and Pb content used in fertilizers and pesticides. This statement was confirmed by [21] and [39] stating that pesticides and herbicides contain lead and arsenic which contaminate the sea and accumulate in sediments and bivalves. Pollutants from antifouling ship paint and ship fuel are associated with Cd and Pb metal contamination [48;5]. Cd and Pb are not detected in water but present in sediments because sediment has the ability to absorb heavy metals better than the water [13]. [41] further stated that much more a little of 1% metal contaminants (and other pollutants) are dissolved in water, while more than 99% are stored in sediments. Heavy metals contained in the waters are then absorbed in particles and accumulated in sediments.

## Heavy Metal Test Results on Blood Cockle Soft Tissue

The results of heavy metal content in soft tissue showed that Hg was not detected in soft tissue shells. Samples taken in February showed that the lowest Cd content was found at Station C with a value of  $0.60~\text{mg kg}^{-1}$  and the highest was at Station D with a value of  $0.66~\text{mg kg}^{-1}$ . Observations taken in March showed the lowest Cd level found at station B with a value of  $0.56~\text{mg kg}^{-1}$  and the highest was at station C with a value of  $0.70~\text{mg kg}^{-1}$ . From the sample taken in in February showed that the lowest Pb content was found at station C with a value of 0.05~mg / kg and the highest was at station D with a value of  $0.1~\text{mg kg}^{-1}$ . In March showed the lowest Pb level found at station D with a value of  $0.06~\text{mg kg}^{-1}$  and the highest was at station A and B with a value of  $0.09~\text{mg kg}^{-1}$  Cd, Pb and Hg test results from soft tissue Blood Cockle are provided in Table 3.

**Table 3.** Heavy Metal Content of A.granosa Soft Tissue

Parameter	Heavy Metal Content of <i>Anadara granosa</i> Soft Tissue at Station (mg.kg <sup>-1</sup> )				Standard SNI* & EC	Month
	A	В	C	D	2006** (mg.kg <sup>-1</sup> )	1,1011011
Cd	0,64 ±0,18	0,64 ±0,20	$0,60 \pm 0,02$	$0.66 \pm 0.08$	1	
Pb	$0.08 \pm 0.03$	$0,08 \pm 0.02$	$0.05 \pm 0$	$0,10\pm0,05$	1,5	February
Hg	Nd	Nd	Nd	Nd	1	
Cd	$0,62 \pm 0,05$	$0,56 \pm 0,01$	$0,70\pm0,06$	$0,66 \pm 0,13$	1	
Pb	$0,09 \pm 0,03$	$0,09 \pm 0,03$	$0,08 \pm 0,04$	$0,06 \pm 0,01$	1,5	March
Hg	Nd	Nd	Nd	Nd	1	

<sup>\*</sup>SNI (National Indonesian Standard number SNI 7387: 2009 \*\*EC Commission Regulation 2006

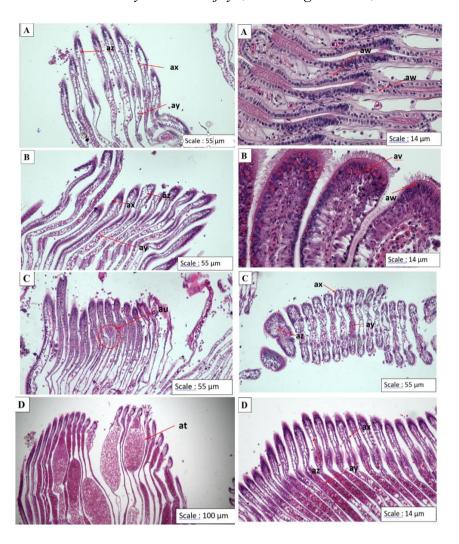
The results of the Kruskal Walis test on the Cd content from the soft tissue of Blood Cockle at the four stations showed there was no difference in Cd content from four stations with p-value of 0.983 in February and 0.421 in March (>0.05). Kruskal Walis test for Pb levels at the four stations gave the p-value of 0.435 in February and 0.487 in March (>0.05) which means that there were no significant differences of Pb content in the four stations.

The value of Cd and Pb content in *A. granosa* soft tissue are still below the maximum level of Cd and Pb content in Indonesian Nasional Standard [28] and the European Union through EC (2006) [8]. Therefore, *A. granosa* from the Wedung Waters are still safe for consumption. Heavy metals enter the marine invertebrates body generally through a solution and food [35]. Heavy metal uptake involves free metal ions such as Cd <sup>++</sup> and Zn <sup>++</sup> as bioavailable dissolved forms, although at equilibrium free metal ions at sea, often only represent a small percentage of the total concentration of dissolved metals [50;43;29]. Absorption of heavy metals into the vertebral body can occur passively through diffusion or with permeable surfaces. Scenarios that allow for passive absorption through a diffusion process are that heavy metals will bind intracellularly to metal-binding ligands [49;44;36].

## Histopathology test results

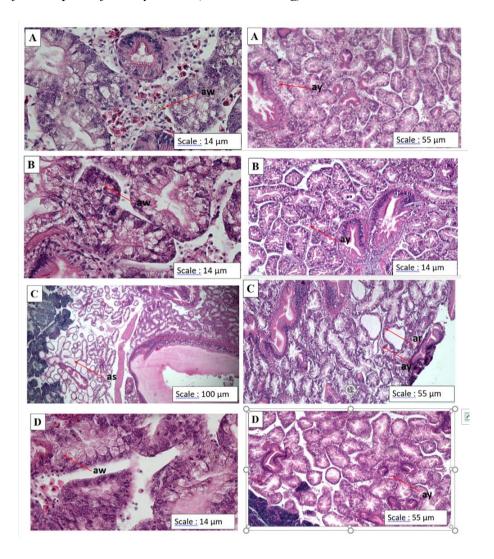
Histopathological observations on the Gills of Blood Cockle in February showed the presence of inflammation as indicated by the presence of hyperplasia cell goblet, cilia degeneration, hemocyte infiltration and pigmentation and the presence of gill swelling (Figure 2). Histopathological observations of gills in March also showed the same results. Gills are known as one of the organs that are most affected in many types of toxicity as it become entrances of toxins dissolved in water [51]. *Cladophora glomerata* contaminated by Cd<sup>++</sup> showed experienced changes in gills such as curling and secondary lamella fusion, epithelial hypertrophy, epithelial hyperplasia, pillar cell damage, edema, swelling, aneurysm, necrosis, and increased mucus secretion. This condition is getting worse as the dose and exposure time increased [31]. Kahn [19] stating the existence of histopathological in *Anodonta cygnea* due to heavy metal contamination with a sequence of levels of metals in soft tissue follows the order of Fe> Zn> Mn> Pb> Cu> Cr> Ni> Cd, indicating an abnormality in the gills such as flat fusion, widening of the hemolymphatic sinus, degeneration of the cilia, hemocyte infiltration, and gill swelling.

The pathological effects of accumulated Cd are thought to negatively interfere with the transport of oxygen needed for respiration by gills [30]. Degeneration of the gills, in general, results in an unbalanced oxygen delivery, which can trigger functional problems in other metabolic organs such as the liver and kidneys [51]. Amin [1] stated the ciliary degeneration in *P. viridis* due to exposure to heavy metals such as Pb that entered during the process of taking food and food diffusion caused a decrease in the growth rate and caused death.



**Figure 2**. Gill histopathology of *A. granosa* (A) Station A, (B) Station B, (C) Station C and (D) Station D.(az) Hyperplasia goblet cell, (ay) Hemocyte infiltration (ax) Cilia degeneration, (aw) Pigmentation, (av)Hemocyte granulocyte,(at) Gill swelling,(au) Lamella fussion.

Histopathological observation of the digestive glands, namely hepatopancreas in four samples also showed an indication of inflammation, it was seen the presence of hemocyte infiltration, pigmentation and necrotic epithelial cells and epithelial cell flattening (Figure. 3). Khan [19] also found similar histopathological effects of Pb and Cd from contaminated areas on *Anodonta cygnea* (Linea, 1876) in the form of inflammation, hydropic vacuoles, and lipofuscin pigments. Sheir and Handy [42] found that *Mytilus edulis* exposed to Cd with concentration of 20  $\mu$ g / 1 and 50  $\mu$ g / 1, the highest concentration of Cd presents with sequence of the digestive glands, gills, and then posterior adductor muscle. Cd causes pathology in the digestive gland, especially at a dose of 20  $\mu$ g / 1 which causes inflammation in the form of hemocyte infiltration.



**Figure 3.** Hepatopancreas histopathology of *A. granosa* A) Station A, (B) Station B, (C) Station C and (D) Station D (ay) Hemocyte infiltration, (aw) Pigmentation, (as) Flattening epithelium cells, (ar) Necrotic epithelium cells

## **Conclusions**

The waters of Wedung Demak are not contaminated by Hg metal. However, Pb and Cd are found in the sediments and soft tissue of *A. granosa*. Cd content in sediment ranges from 0.08 to 0.2 mg. Kg<sup>-1</sup> while Pb ranges from 3.35 to 6.46 mg. Kg<sup>-1</sup>. The value is still not too dangerous for the environment. In soft tissue of Blood Cockle, Cd found around 0.56 - 0.7 mg.kg<sup>-1</sup> and Pb of 0.05 - 0.1 mg.kg<sup>-1</sup> which is still safe for consumption. Histopathology test result indicate inflammation of the gills and hepatopancreas.

#### Reference

- [1] Amin, B., E. Afriyani dan M.A. Saputra. 2011. Distribusi Spasial Logam Pb dan Cu pada Sedimen dan Air Laut Permukaan di Perairan Tanjung Buton Kabupaten Siak Provinsi Riau. Jurnal Teknobiologi II (1): 1–8.
- [2] Ansari, T. M., Marr, I. L., & Tariq, N. (2004). Heavy metals in marine pollution perspective—a mini review. *Journal of Applied Sciences*, 4(1), 1-20.
- [3] Broom, M. J. (Ed.). (1985). *The biology and culture of marine bivalve molluscs of the genus Anadara* (Vol. 12). WorldFish.
- [4] Brotohadikusomo, N. A. (1994). The ecology of two species of blood clams, Anadara granosa (L.) and Anadara antiquata (L.) in Central Java, Indonesia (Doctoral dissertation, Prifysgol Bangor University).
- [5] Chakraborty, S., Bhattacharya, T., Singh, G., & Maity, J. P. (2014). Benthic macroalgae as biological indicators of heavy metal pollution in the marine environments: A biomonitoring approach for pollution assessment. *Ecotoxicology and Environmental Safety*, 100, 61-68.
- [6] Costa, P. M. (2017). The handbook of histopathological practices in aquatic environments: Guide to histology for environmental toxicology. Academic Press.
- [7] Dabwan, A. H., & Taufiq, M. (2016). Bivalves as bio-indicators for heavy metals detection in Kuala Kemaman, Terengganu, Malaysia. *Indian journal of science and technology*, 9(9), 1-6.
- [8] EC (2006) Commission regulation (EC) No 188/2006. Off J Eur Un Legis 49(31):10–11.
- [9] Farejiya, M. K., & Dikshit, A. K. (2016). Assessment of Heavy Metal Concentrations in Tunas Caught from Lakshweep Islands, India. *World Academy of Science, Engineering and Technology, International Journal of Environmental, Chemical, Ecological, Geological and Geophysical Engineering*, 10(6), 697-700.
- [10] Gandaseca, S., Wahab, N. L. A., Pazi, A. M. M., Rosli, N., & Zaki, P. H. (2016). Comparison of Water Quality Status of Disturbed and Undisturbed Mangrove Forest at Awat-Awat Lawas Sarawak.
- [11] Gao, X., & Chen, C. T. A. (2012). Heavy metal pollution status in surface sediments of the coastal Bohai Bay. *Water research*, 46(6), 1901-1911.
- [12] Gimeno-García, E., Andreu, V., & Boluda, R. (1996). Heavy metals incidence in the application of inorganic fertilizers and pesticides to rice farming soils. *Environmental pollution*, 92(1), 19-25.
- [13] Goldsmith, S. L., Krom, M. D., Sandler, A., & Herut, B. (2001). Spatial trends in the chemical composition of sediments on the continental shelf and slope off the Mediterranean coast of Israel. *Continental Shelf Research*, 21(16-17), 1879-1900.
- [14] González-Fernández, D., Garrido-Pérez, M. C., Nebot-Sanz, E., & Sales-Márquez, D. (2011). Source and fate of heavy metals in marine sediments from a

- semi-enclosed deep embayment subjected to severe anthropogenic activities. *Water, Air, & Soil Pollution, 221*(1-4), 191.
- [15] IADC/CEDA. 1997. Convention, Codes, and Conditions: Marine Disposal. Environmental Aspects of Dredging 2a. 71 hal.
- [16] Islam, M. S., Ahmed, M. K., Raknuzzaman, M., Habibullah-Al-Mamun, M., & Islam, M. K. (2015). Heavy metal pollution in surface water and sediment: a preliminary assessment of an urban river in a developing country. *Ecological indicators*, 48, 282-291.
- [17] Katalay, S., Yavasoglu, A., Yigitturk, G., Oltulu, F., Sari, G., Yavasoglu, N. U. K., & Karabay, U. (2016). Histological effects of pollution on gill and hepatopancreas tissues of black mussels (*M. galloprovincialis L.*) from Izmir Bay of Turkey. *Fresenius Environmental Bulletin*, 25(5), 1461-1467.
- [18] Kementerian Kelautan dan Perikanan, "Kelautan dan perikanan dalam angka tahun 2015." (2015).
- [19] Khan, M. I., Khisroon, M., Khan, A., Gulfam, N., Siraj, M., Zaidi, F., & Qadir, F. (2018). Bioaccumulation of heavy metals in water, sediments, and tissues and their histopathological effects on *Anodonta cygnea* (Linea, 1876) in Kabul River, Khyber Pakhtunkhwa, Pakistan. *BioMed research international*, 2018.
- [20] Li, J., Sun, C., Zheng, L., Jiang, F., Wang, S., Zhuang, Z., & Wang, X. (2017). Determination of trace metals and analysis of arsenic species in tropical marine fishes from Spratly islands. *Marine pollution bulletin*, 122(1-2), 464-469.
- [21] Lias, K., Jamil, T., & Aliaa, S. N. (2013). A preliminary study on heavy metal concentration in the marine bivalves *Marcia marmorata* species and sediments collected from the coastal area of Kuala Perlis, North of Malaysia. *IOSR journal of applied chemistry*, 4(1), 48-54.
- [22] Martín-Díaz, M. L., Blasco, J., de Canales, M. G., Sales, D., & DelValls, T. Á. (2005). Bioaccumulation and toxicity of dissolved heavy metals from the Guadalquivir Estuary after the Aznalcollar mining spill using *Ruditapes philippinarum*. *Archives of environmental contamination and toxicology*, 48(2), 233-241.
- [23] Nasional, Badan Standardisasi. "SNI 2354.5: 2011." Tentang Penentuan Kadar Logam Berat Timbal (Pb) dan Kadmium (Cd) pada Produk Perikanan. Badan Standarisasi Nasional, Jakarta (2011).
- [24] Nasional, Badan Standardisasi. "SNI 2354.6:2016" Cara uji kimia Bagian 6: Penentuan Kadar Logam Berat Merkuri (Hg) Pada Produk Perikanan. *Badan Standarisasi Nasional, Jakarta* (2016).
- [25] Nasional, Badan Standarisasi. "SNI 6989. 16: 2009." *Air dan Air limbah Cara Uji Kadmium (Cd) secara Spektrofotometri Serapan Atom (SSA)—nyala* (2009).
- [26] Nasional, Badan Standarisasi. "SNI 6989. 78: 2011." *Air dan Air limbah Cara Uji Raksa (Hg) secara Spektrofotometri Serapan Atom (SSA)–nyala* (2009).

- [27] Nasional, Badan Standarisasi. "SNI 6989. 8: 2004." *Air dan Air limbah Cara Uji Timbal (Pb) secara Spektrofotometri Serapan Atom (SSA)–nyala* (2004).
- [28] Nasional, Badan Standarisasi. "SNI 7387: 2009." Tentang Batas Maksimum Cemaran Logam Berat dalam Pangan. Jakarta: Badan Standarisasi Nasional (2009).
- [29] Nugegoda, D., & Rainbow, P. S. (1988). Effect of a chelating agent (EDTA) on zinc uptake and regulation by *Palaemon elegans* (Crustacea: Decapoda). *Journal of the Marine Biological Association of the United Kingdom*, 68(1), 25-40.
- [30] Omer, S. A., Elobeid, M. A., Fouad, D., Daghestani, M. H., Al-Olayan, E. M., Elamin, M. H., & El-Mahassna, A. (2012). Cadmium bioaccumulation and toxicity in tilapia fish (*Oreochromis niloticus*). *Journal of animal and veterinary advances*, *11*(10), 1601-1606.
- [31] OTLUDİL, B., AKIN, H. K., & Erhan, Ü. N. L. Ü. (2017). Effects of sub-lethal exposure of cadmium on histopathology of gills of Nile tilapia, *Oreochromis niloticus* and the mitigating effects of *Cladophora glomerata*. *Türk Biyoloji Dergisi*, 30(1), 24-30.
- [32] Poleksic, V., Lenhardt, M., Jaric, I., Djordjevic, D., Gacic, Z., Cvijanovic, G., & Raskovic, B. (2010). Liver, gills, and skin histopathology and heavy metal content of the Danube sterlet (*Acipenser ruthenus* Linnaeus, 1758). *Environmental Toxicology and Chemistry: An International Journal*, 29(3), 515-521.
- [33] Qian, Y., Zheng, M. H., Gao, L., Zhang, B., Liu, W., Jiao, W., & Xiao, K. (2005). Heavy metal contamination and its environmental risk assessment in surface sediments from Lake Dongting, People's Republic of China. *Bulletin of environmental contamination and toxicology*, 75(1), 204-210.
- [34] Azizah, R., Malau, R., Susanto, A. B., Santosa, G. W., Hartati, R., Irwani, I., & Suryono, S. (2018). Kandungan Timbal Pada Air, Sedimen, Dan Rumput Laut *Sargassum sp.* Di Perairan Teluk Awur, Jepara. *Jurnal Kelautan Tropis*, 21(2), 155-156.
- [35] Rainbow, P. S. (1990). Heavy metal levels in marine invertebrates. *Heavy metals in the marine environment*, 67-79.
- [36] Rainbow, P. S., (1988), The significance of trace metal concentrations in decapods, in *Symp. Zool. Soc. London, 59, Aspects of Decapod Crustacean Biology,* Fincham, A. A. and Rainbow, P. S., Eds., Oxford University Press, Oxford, 291.
- [37] Rodriguez-Iruretagoiena, A., Rementeria, A., Zaldibar, B., de Vallejuelo, S. F. O., Gredilla, A., Arana, G., & de Diego, A. (2016). Is there a direct relationship between stress biomarkers in oysters and the amount of metals in the sediments where they inhabit? *Marine pollution bulletin*, 111(1-2), 95-105.
- [38] Rosioru, D. M., Oros, A., & Lazar, L. (2016). Assessment of the heavy metals contamination in bivalve *Mytilus galloprovincialis* using accumulation

- factors. Journal of Environmental Protection and Ecology, 17(3), 874-884.
- [39] Rumsby, A. (2011). Significance of arsenic in sediments of Lake Rotoroa (Hamilton Lake). *Waikato Regional Council Technical Report*, 18.
- [40] Salam, M. A., Paul, S. C., Noor, S. N. B. M., Siddiqua, S. A., Aka, T. D., Wahab, R., & Aweng, E. R. (2019). Contamination profile of heavy metals in marine fish and shellfish. *Global Journal of Environmental Science and Management*, *5*(2), 225-236.
- [41] Salomons, W., & Stigliani, W. M. (Eds.). (2012). Biogeodynamics of pollutants in soils and sediments: risk assessment of delayed and non-linear responses. Springer Science & Business Media.
- [42] Sheir, S. K., & Handy, R. D. (2010). Tissue injury and cellular immune responses to cadmium chloride exposure in the common mussel *Mytilus edulis*: modulation by lipopolysaccharide. *Archives of environmental contamination and toxicology*, 59(4), 602-613.
- [43] Simkiss, K. (1983). Lipid solubility of heavy metals in saline solutions. *Journal of the marine biological Association of the United Kingdom*, 63(1), 1-7.
- [44] Simkiss, K. (1982). Metal detoxification and bioaccumulation in molluscs. *Mar. Biol. Lett.*, *3*, 187.
- [45] Stankovic, S., & Jovic, M. (2012). Health risks of heavy metals in the mediterranean mussels as seafood. *Environmental chemistry letters*, 10(2), 119-130.
- [46] Statistik, Badan Pusat, (2017). Kecamatan Wedung dalam angka, Badan Pusat Statistik Kabupaten Demak.
- [47] Tjahjono, A., & Suwarno, D. (2018). The Spatial Distribution of Heavy Metal Lead and Cadmium Pollution and Coliform Abundance of Waters and Surface Sediment in Demak. *Journal of Ecological Engineering*, 19(4).
- [48] Turner, A. (2010). Marine pollution from antifouling paint particles. *Marine Pollution Bulletin*, 60(2), 159-171.
- [49] Williams, R. J. P. (1981). Physico-chemical aspects of inorganic element transfer through membranes. *Philosophical Transactions of the Royal Society of London. B, Biological Sciences*, 294(1071), 57-74.
- [50] Wright, D. A. (1977). The effect of salinity on cadmium uptake by the tissues of the shore crab *Carcinus maenas*. *Journal of Experimental Biology*, 67(1), 137-146.
- [51] Yılmaz, M., Ersan, Y., Koç, E., Özen, H., & Karaman, M. (2011). Toxic Effects of Cadmium Sulphate on Tissue Histopathology and Serum Protein Expression in European chub, *Leuciscus cephalus* (Linnaeus, 1758).